

# Can EDTA Change MRSA into MSSA? A Future Prospective!

SONIA JAIN<sup>1</sup>, MANIDEEPA SENGUPTA<sup>2</sup>, SOMA SARKAR<sup>3</sup>, SOUGATA GHOSH<sup>4</sup>, ANITA NANDI (MITRA)<sup>5</sup>, ANURADHA SINHA<sup>6</sup>, SRIPARNA CHAKRAVORTY<sup>7</sup>

## ABSTRACT

**Introduction:** In the present era we are left behind with limited options for the treatment of serious infections caused by multidrug resistant *S.aureus*, most remarkably nosocomially acquired Methicillin resistant *S.aureus* (MRSA). The problem increases more when these strains easily become multidrug resistant (MDR) due to biofilm formation. Those *staphylococcal* species that are vancomycin and linezolid resistant are also resistant to other antistaphylococcal agents which call for an urgent intervention to develop newer antimicrobial agents.

**Aim:** The present study was undertaken with the aim to evaluate the antibiofilm effect of EDTA against the biofilm forming MRSA isolates, isolated from different clinical infections.

**Materials and Methods:** The biofilms formed on polystyrene microtitre plates by the MRSA strains were treated by different concentrations of EDTA to find out its anti-biofilm activity. Further simultaneously the antibiotic susceptibility pattern was noted down to check whether the MRSA strains become MSSA (Methicillin sensitive *S.aureus*).

**Results:** Our data demonstrates that EDTA at 4mM concentration inhibits biofilm of MRSA and at 20 mM have an ability to reduce and dissociate the biofilm membrane, allowing the antibiotics to enter and convert MRSA strains into MSSA.

**Conclusion:** These findings suggest that commercially available EDTA could be used in future to control MRSA and its biofilm-related infections.

**Keywords:** Clinical isolates, Strong biofilm formers, Antibiofilm effect

## INTRODUCTION

*Staphylococcus aureus* causes multiple infections, ranging from infections of skin and soft-tissue, foreign-body infections, osteomyelitis, pneumonia, septic arthritis to life threatening infections like endocarditis and septicaemia [1] as well as a leading cause of infections of implants and indwelling medical device [2] both in hospital and community settings. Methicillin-resistant *Staphylococcus aureus* (MRSA), was first reported in 1961 and thereafter it has become a major nosocomial pathogen which is prevalent all over the world [3]. Treatment due to infections caused by MRSA strains are becoming more critical and costly compared to those due to MSSA. The capacity to produce biofilm by MRSA strains, further contributes to antimicrobial resistance thereby increasing the rate of morbidity and mortality [4]. According to the Centers for Disease Control (CDC) and the National Institutes of Health (NIHS), 65% to 80% of all chronic infections are caused by the microorganisms producing biofilm [5,6]. Biofilm is defined as a population of microorganisms adhered irreversibly on a biotic or abiotic surface, and enclosed in a self-synthesized hydrated matrix of protein, polysaccharides, exopolymeric substances (EPS) and nucleic acids. It also involves attachment and immobilization, cell to cell adhesion and interaction, micro-colony formation with confluent 3-D structure [4,7].

With the increase in biofilm-related infection, researchers have tried to examine new ways to prevent and remove microbial biofilms from an infected site to control the surface attached bacteria. Among the various strategies have been proposed to control biofilm, one of them being using “nonantibiotic drugs” for their antimicrobial or antibiofilm activities [8]. One of the such examples is coating various medical devices with EDTA aimed at inhibiting initial cell adhesions [9].

Compounds like sodium ethylenediaminetetraacetate (EDTA) possess some antimicrobial activity, the potency of which can be increased by combining other molecules [10] or antimicrobials along with EDTA [11,12]. It should be mentioned here that cations are required to stabilize the negatively charged polysaccharides which is

required to hold the biofilm together EDTA forms strong complexes with these cations thereby chelating them and destabilizing the formed biofilm [13]. In the present study, we have tried to find out the inhibitory effect of EDTA against MRSA strains with a capacity to form the biofilm.

## MATERIALS AND METHODS

**The clinical isolates of MRSA and its microbiological testing:** After getting ethical clearance from the institutional Ethical Committee of Medical College and Hospital, Kolkata quasi Experimental (nonrandomized, pre-post-intervention) study was performed on the 25 strong biofilm forming clinical strains isolated in the Microbiology Department, Medical College, Kolkata from Jan 2013 to Dec 2013. Firstly, *Staphylococcus aureus* strains were isolated from the different samples (pus, sputum, urine, blood, fluid, and other miscellaneous samples like central venous catheter tip, endotracheal tube, orthopedic implants, etc) of the patients received by the Microbiology Department for routine examination. The samples were initially streaked in Blood agar plates followed by 24 hours incubation at 37°C. *Staphylococcus aureus* was confirmed by Gram staining, slide coagulase test, tube coagulase test and anaerobic mannitol fermentation by using standard methods [14]. All the confirmed *Staphylococcus aureus* strains were further screened for methicillin resistance with the help of cefoxitin discs (30 µg/disc) (Hi-Media, India) by using Kirby-Bauer disc diffusion method. The isolates were considered resistant if the zone of inhibition was 21mm or less (10). Followed by other antibiotics like Clindamycin (2µg) Erythromycin (15 µg), Vancomycin (30 µg), Amikacin (30µg), Gentamycin (10µg), Cefuroxime (µg), Levofloxacin (5µg), Ciprofloxacin (5 µg), Linezolid (30 µg), Doxycycline (30 µg), Amoxicillin (10µg), Amoxyclav (30µg) on Mueller-Hinton agar. These antibiotic discs were obtained from commercial source. Results were interpreted according to CLSI guidelines (2014) [15]. Throughout the study *S. aureus* ATCC 25923 was used as a control.

**Bacterial DNA isolation:** Genomic DNA was isolated with phenol and chloroform method and was precipitated with ethanol by

standard methods [16]. The only addition was lysostaphin (40 mg per ml) and incubation for 30 min at 37°C prior to addition of RNase. The DNA was recovered by centrifugation, vacuum dried and finally dissolved in 200µl Tris- EDTA buffer (10mM Tris, 1mM EDTA (pH 8.0), and concentration was determined spectrophotometrically at 260nm. An amount of 10ng template DNA was used in this study.

**PCR primer design and amplification:** By using specific accession numbers nucleotide sequences, each of the desired genes were obtained from GeneBank. The identities of *S. aureus* isolates were confirmed by species-specific gene amplification (16S rRNA) and the presence of *mecA* gene, responsible for methicillin resistance was carried out by employing the sets of gene specific primers in PCR assay. For the PCR amplification, the reaction mixture composition was similar for both the genes and oligonucleotide primers used in the study. DNA amplification was performed in 25µl of reaction mixture that contained 2.5 µl 10x buffer, 2.5 µl of 200mM dNTP mix (I.D.T), 2 µl of 25 mM of MgCl<sub>2</sub>, 1.5µl each of a pair of primers forward and reverse (10 pmol/µl, I.D.T) , 10 ng of *staphylococcal* template DNA and 1 U of Ampli-TechgoldTaq DNA polymerase. PCR amplification was carried out in a DNA thermal cycler (Applied Biosystem 2720) under reaction conditions as described. Denaturation, annealing and extension temperatures were kept at 94°C, 64 °C (for 16S r RNA), 54 °C (for *mecA*) and 72°C respectively for each gene amplification [17,18].

The amplification product was resolved by electrophoresis on 1.5 % agarose gel at 70V (constant voltage) and visualized with ultraviolet light after staining with ethidium bromide. Product size was determined by using the 500bp DNA molecular weight ladder (NEB).

**Biofilm formation and its treatment:** For detection of biofilm we used quantitative microtitre plate method [19]. Isolated colonies of MRSA strains were inoculated in 5ml of trypticase soy broth (TSB) with 1% glucose and then it was incubated at 37°C for 24 h. The cultures were then diluted with fresh medium at ratio of 1:100. Each well of sterile 96 well-flat polystyrene tissue culture treated plates (Tarson, India) was inoculated with 200 µL of the diluted broth cultures of different MRSA strains isolated from various samples. *S. aureus* ATCC 25923 was used as positive biofilm control. Negative control wells were inoculated with sterile broth. The plates were incubated at 37°C for 24 h. After incubation, the wells were washed with 0.2 ml of phosphate buffer saline (PBS) (pH 7.2) four times. This removed free floating bacteria. Biofilm formed by bacteria adherent to the wells were fixed by keeping in hot air oven at 120°C for 60 minutes and were then stained by crystal violet (CV) (0.1%). Excess stain was removed by using deionized water by rinsing four times with sterile distilled water and subsequent decolorizing with 30% acetic acid. Optical density (OD) of stained adherent biofilm was obtained by using micro ELISA autoreader (model 680, Biorad, UK) at wavelength 570 nm. Uninoculated wells containing TSB were used as blanks. Blank-corrected absorbance values were used for reporting biofilm production. The test was performed in triplicate and repeated thrice [Table/Fig-1a]. The biofilm formation was interpreted according to the criteria laid down by Stepanovic et al., [20].

< 0.120 Non Biofilm producer.

0.120 – 0.240 Moderate Biofilm producer.

> 0.240 Strong Biofilm producer.

**Inhibition by EDTA treatment:** The method of Amalaradjou et al., was used to assess the ability of the EDTA to inhibit MRSA biofilm production [21]. MRSA strains were separately grown overnight in TSB at 37°C.

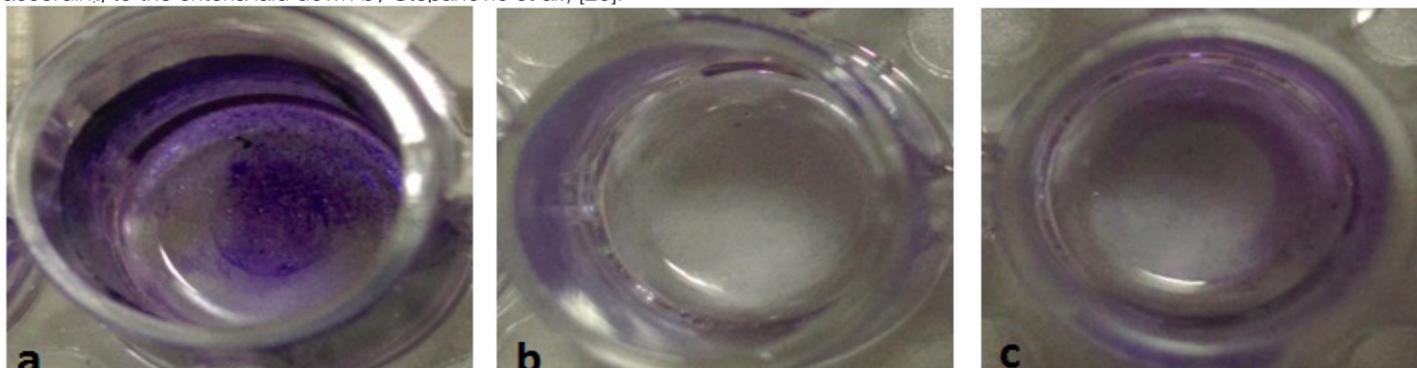
Two hundred microliters of this culture were used as the inoculum (~6.0 log CFU) and transferred into sterile 96-well polystyrene tissue culture plates, followed by the addition of 0 (negative control), 0.5 (0.2 µl), 1 (0.4 µl), 2 (0.8 µl) and 4(1.6 µl) mM of EDTA. The plates were incubated at 37°C for 24 hrs and washed four times with 200 µL of sterile PBS, dried and stained with 1% crystal violet for ELISA reading. After rinsing, optical density (OD) of stained adherent biofilm was obtained by using micro ELISA auto reader at wavelength 570 nm. Wells containing TSB without EDTA were used as blank. Triplicate samples were included for each treatment, and the experiment was replicated three times.

**Reduction of biofilm by EDTA treatment:** To evaluate the ability of the EDTA to dissociate MRSA biofilm 200µL of the each bacterial cell suspension (~6.0 logCFU) were inoculated in sterile 96-well polystyrene tissue culture plates and incubated at 37°C for 24 h without agitation for biofilm production [20]. The overnight aggregated culture (formed biofilm) was then exposed for 24 hours with different concentrations 0 (negative control), 2.5 (1 µl), 5 (2µl), 10 (4 µl) and 20 (8 µl) mM of EDTA by adding it to the microtitre plate. Thereafter, the wells were washed four times with 200 µL of sterile PBS, dried and stained with 1% crystal violet for ELISA reading. After rinsing, the absorbance of the adherent biofilm was measured at 570 nm in a microplate reader. Wells containing TSB without EDTA were used as control. Triplicate wells were included for each treatment.

**Determination of Bacterial Counts and Sensitivity pattern in Biofilm:** The antibiofilm effect of EDTA was studied by counting surviving bacterial populations in the biofilm by viable plate count method [21]. Following treatment with EDTA, the wells were washed four times with PBS, and the adherent biofilm was scraped and plated on Nutrient Agar. The plates were incubated at 37°C for 24 h before enumerating the bacterial colonies. The isolated bacterial colonies were then subsequently tested for methicillin and other antibiotic resistance by Kirby-Bauer disc diffusion method discs on Mueller-Hinton agar. The plates were incubated at 37°C for 24 hours and zone size was recorded. Minimum Inhibitory Concentration (MIC) for methicillin was also determined by using Ezy MIC strip according to the instructions given by the manufacturer (Hi Media Laboratories, India).

## STATISTICAL ANALYSIS

The t-test was utilized for calculating the difference of mean between OD value for control (0mM EDTA) and different concentration of EDTA used for both reduction and inhibition. The results of biofilm



**[Table/Fig-1a-c]:** Biofilm reduction and inhibition assay. (a. pre-formed biofilm) MRSA biofilms were developed on microtitre plates for 24 hrs, (b. biofilm inhibition) after addition of different concentrations of EDTA and (c. biofilm reduction) after treatment of biofilm with different concentrations of EDTA. Biofilms were rinsed and stained with CV.

inhibition and reduction were significantly lower ( $p < 0.00$ ) for all the different concentrations of EDTA.

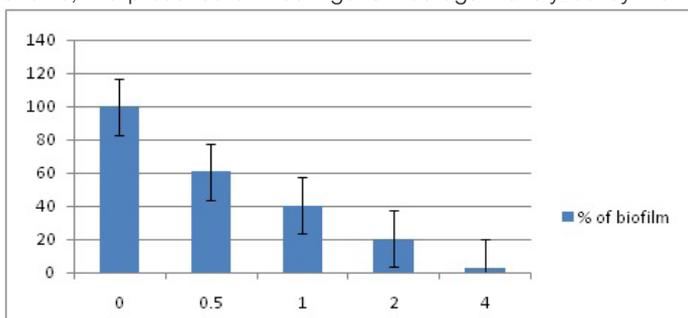
## RESULTS

Samples were collected in the Microbiology Department of Medical College from patients coming to the hospital during the year 2013 in West Bengal, India. Gram positive bacterial pathogen was identified in 2987 samples of which 420 (14.06%) strains were that of *Staphylococcus aureus*. Out of which 96 (22.8 %) were MRSA isolates. From this 25 (26.04%) were strong biofilm formers.

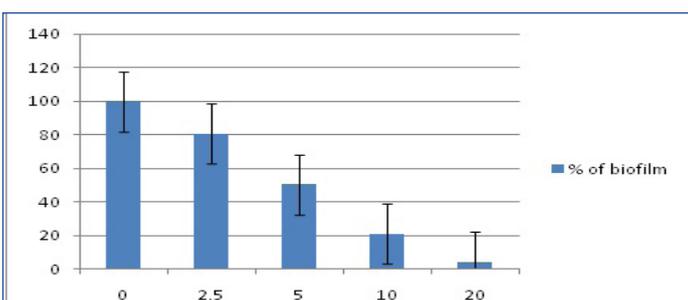
**Biofilm Inhibition:** To assess the ability of EDTA to inhibit biofilm, 25 MRSA samples were loaded in triplicate in 96-well plates in presence of different concentrations of EDTA i.e. 0, 0.5, 1, 2 and 4mM. As seen by crystal violet biofilm quantification at 570nm, significant inhibition (96.96%) was observed in the wells with 4mM concentration for each of 25 MRSA samples [Table/Fig-1b,2].

**Biofilm Reduction:** EDTA was also effective at removing fully formed biofilms of MRSA on polystyrene plates. The 96-well microtitre plates with fully formed biofilm in 25 MRSA samples was treated with different concentrations i.e. 0 (Control), 2.5, 5, 10, 20 mM of EDTA. As seen by crystal violet biofilm quantification at 570nm, significant reduction of biofilm was observed in the wells for each of 25 MRSA samples. EDTA at 20 mM concentration for 24 hrs of exposure, showed almost complete reduction of biofilm (95.5%) of the MRSA strains [Table/Fig-1c,3].

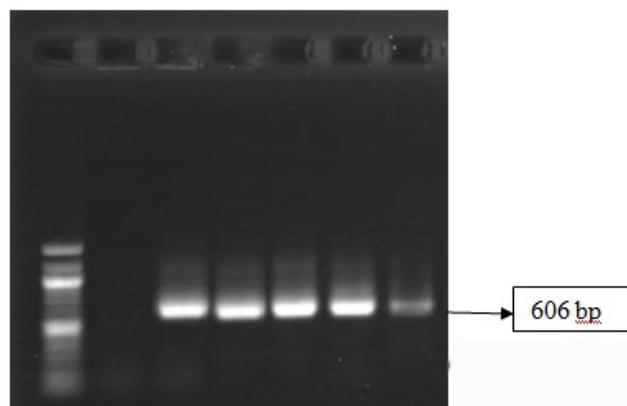
**Conversion of MRSA into MSSA:** The simple viable plate count method helped to demonstrate the surviving bacterial populations in the biofilm. From these bacterial populations, an inoculum was prepared for antimicrobial susceptibility test by Kirby Bauer Disc Diffusion Method. After treatment with EDTA, the MRSA strains which were resistant to all the drugs as mentioned above became sensitive to those drugs including ceftioxin. EDTA, thus converted MRSA strains to MSSA strains. MIC for ceftioxin was again determined by using Ezy MIC strip according to the instructions given by the manufacturer. MIC was found to be  $\leq 22\mu\text{g/mL}$  confirming the conversion of MRSA isolates to MSSA, after treatment with 10mM concentration of EDTA. Further to test whether any genetic change was introduced in EDTA treated MRSA strains, the presence of *mecA* gene was again analyzed by PCR.



**[Table/Fig-2]:** Quantification of biofilm inhibition formed on microtitre plate after treating with different concentrations (mM) of EDTA in each of 25 MRSA samples as shown in Fig. Each experiment was carried out three times, yielding almost similar results. Data are represented as the mean  $\pm$  SEM.



**[Table/Fig-3]:** Reduction of biofilm on microtitre plate after treating with different concentrations (mM) of EDTA in each of 25 MRSA samples. Each experiment was carried out three times, yielding almost similar results. Data are represented as the mean  $\pm$  SEM.



**[Table/Fig-4]:** Agarose gel electrophoresis of PCR-amplified *mecA* genes at 606 bp of EDTA treated MRSA isolates (i) Lane 1: 1000 bp ladder; Lane 2: Negative Control; Lane 6: Positive Control *S. aureus* ATCC 33591 (ii) Lane 3-5: MRSA isolate after EDTA treatment at 2.5, 5, 10 and 20 mM conc

But the PCR amplification revealed the presence of *mecA* genes in genomic isolates, confirming only the phenotypic changes [Table/Fig-4].

## DISCUSSION

Sometimes, the various biomedical devices used for different clinical procedures may be associated with infections due to microbial colonization and multiplication [22]. Among them, *Staphylococci* mainly causes multiple infections on foreign devices like orthopedic and surgical implants, stents, intravenous catheters, infusion pumps, mechanical heart valves, pacemakers, surgical implants etc where biofilms can result in serious complications leading to high morbidity, mortality, and medical costs [23]. For eradicating the biofilm from the infected site it usually requires surgical intervention with antimicrobial therapy [24]. The bacteria enclosed in a biofilm layer are usually multi-drug resistant compare to free floating form. Therefore, a possible therapeutic approach to biofilm associated medical device infections involves the use of EPS-degrading agents that might reduce or remove cell-to-cell and cell-to-surface associations within the biofilm, thus releasing the planktonic cells into the environment and allowing the host immune response and antimicrobial agents to act easily upon them [25,26]. In this study, we examined the potential use of EDTA as a biofilm control agent against MRSA. It has already been reported in some of the literature that EDTA, a chelator of calcium and magnesium, and an anticoagulant, possesses potential activity against planktonic microbial cells and inhibits their biofilms [27,28]. Root et al., also demonstrated the effect of EDTA on biofilm formed by *S. epidermidis* (invitro) on a Hickman catheter made of silicone material. They concluded that a high concentration of EDTA i.e. 108 mmol/l could eradicate the *staphylococcal* biofilm [28]. In one of the studies, EDTA treatment with 40 mg/ml for 24 hrs significantly reduced the viability of biofilms of *S. epidermidis*, MRSA, *P. aeruginosa*, *K. pneumoniae* or *C. albicans* [29]. On the other hand, according to some authors higher than 2 mmol/l concentrations of EDTA, was in vitro toxic for viability of microbial cell cultures [30]. In another study conducted by Marek et al., EDTA inhibited formation of biofilm in *S. epidermidis* on medical devices at 1.0–2.0 mmol/l concentration and reduced the mature biofilm in some strains at 2.0–4.0 mmol/l EDTA whereas for the other isolates, EDTA (>32 mmol/l) was required at higher concentrations [31]. But one of the studies showed no significant decrease in *Staphylococcus* biofilm, even after an exposure to the EDTA for 60 min [32]. EDTA may also be combined with other molecules like minocycline, ovotransferin, protamine sulfate or antibiotics for a synergistic antibiofilm effect against *Staphylococcus* spp [33]. Another recent study indicated that more than 1.5 mg/ml of EDTA inhibited the growth of *L. monocytogenes*. However, when EDTA was combined with ovotransferrin, the strength of its action decreased [34]. Our results showed that EDTA at 4mM and 20 mM concentration for 24 hours, demonstrates a significant inhibition and

reduction of biofilm of the MRSA strains respectively. EDTA has been shown to lead to phenotypic mutation by converting MRSA into MSSA. This could be probably due to the fact that EDTA selectively solubilizes EPS membrane and increases membrane permeability of MRSA, allowing the antibiotics to penetrate inside thereby reducing methicillin resistance. Supporting this there are some reports stating that the inhibitory effect of EDTA on *staphylococcal* biofilm formation is due to the fact that EDTA causes the chelation of metal ions [33]. The appropriate biofilm surface properties are determined by the presence of PIA (Polysaccharide Intracellular Antigen) which in turn depends on the Mg<sup>2+</sup> ions level [35]. Therefore, the decreased level of Mg<sup>2+</sup> ions (which is mainly due to chelation by EDTA) inhibits the adhesion process, thereby inhibiting the biofilm formation [33].

Since MRSA have the ability to persist in the hospital environment and form biofilms on a wide variety of biotic and abiotic surfaces, EDTA can be used as potential therapeutic and prophylactic antimicrobial agent. With the help of aseptic techniques and proper utilization of EDTA solutions, foreign body associated infections can be reduced, thus decreasing the morbidity and mortality rate.

## CONCLUSION

In conclusion, our study demonstrates that EDTA is effective in preventing biofilm formation by MRSA and inactivating pre-formed biofilms on polystyrene plates. These results suggest that EDTA can be potentially used as a sanitizer for hospital surfaces. However, further experiments are needed to evaluate the efficacy of EDTA in comparison with other anti-MRSA therapies both invitro and in vivo. In future, these findings would be beneficial in our battle against superbugs.

**Conflict of Interests:** The authors declare that there is no conflict of interests regarding the publication of this paper.

## ACKNOWLEDGMENT

We acknowledge Dr. Ranjan Nandy, (Scientist E, NICED) for allowing us to carry out the work in his laboratory and for expert technical assistance regarding the PCR, etc. We are also grateful to Dr. Mousumi Datta, Assistant Professor, Department of Community Medicine, Medical College, Kolkata for helping us in statistical calculations. This study was self supported.

## REFERENCES

- [1] Lowy FD. *Staphylococcus aureus* infections. *N Engl J Med*. 1998;33(9):520-32.
- [2] Donlan RM. Biofilms and device-associated infections. *Emerg Infect Dis*. 2001;7:277-81.
- [3] Jevons MP. Celbenin-resistant *staphylococci*. *BMJ*. 1961;1:124-25.
- [4] Otto M. *Staphylococcal* biofilms. *Curr Top Microbiol Immun*. 2008;32(2):207-28.
- [5] Potera C. Forging a link between biofilms and disease. *Science*. 1999;28(3):1837-39.
- [6] NIH research on microbial biofilms. <http://grants.nih.gov/grants/guide/pa-files/PA-03-047.html>.
- [7] Gotz F. *Staphylococcus* and biofilms. *MolMicrobiol*. 2002;43:1367-78.
- [8] Cederlund H, Mardh PA. Antibacterial activities of non-antibiotic drugs. *J Antimicrob Chemother*. 1993;3(2):355-65.
- [9] Reardon DM, Warner B, Trowbridge A. EDTA, the traditional anticoagulant of haematology: with increased automation is it time for a review? *Med Lab Sci*. 1991;4(8):72-75.
- [10] Branan JK, Davidson PM. Enhancement of nisin, lysozyme, and monolaurin antimicrobial activities by ethylenediaminetetraacetic acid and lactoferrin. *Int J Food Microbiol*. 2004;9(0):63-74.
- [11] Carneiro D, Melo MAS, Cassar CL, Miles RJ. Trisodium phosphate increase sensitivity of Gram-negative bacteria to lysozyme and nisin. *J Food Prot*. 1998;6(1):839-44.
- [12] Gill AO, Holley RA. Interactive inhibition of meat spoilage and pathogenic bacteria by lysozyme, nisin and EDTA in the presence of nitrite and sodium chloride at 24 °C. *Int J Food Microbiol*. 2003;8(0):251-59.
- [13] Kala R, Chauhan H, Rajput A, Kuttu R. Biofilm characterization and quorum quenching in pathogenic strains *Staphylococcus aureus* and *Pseudomonas aeruginosa* *International Journal of Advanced Biotechnology and Research*. 2012;3:515-22.
- [14] Mackie and Mccartney Practical Medical Microbiology 14<sup>th</sup> Edition, *Staphylococcus*: Cluster-forming Gram-positive cocci; 11: 245-261.
- [15] Clinical and Laboratory Standards Institute. Performance standards for antimicrobial disk susceptibility tests, Twenty-fourth Informational Supplement (M100 – S24), 2014.
- [16] Sambrook J, Fritsch EF, Maniatis T. *Molecular Cloning: A Laboratory Manual*. 2<sup>nd</sup> edition. New York: Cold Spring Harbor Laboratory Press, 1989
- [17] Monday S, Bohach G. Use of Multiplex PCR To Detect Classical and Newly Described Pyrogenic Toxin Genes in *Staphylococcal* Isolates. *Journal of clinical microbiology*. 1999;3:3411-14.
- [18] Khan AU, Sultan A, Tyagi A, Zahoor S, Akram M, Kaur S, et al. Amplification of *mecA* gene in multi-drug resistant *Staphylococcus aureus* strains from hospital personnel. *J Infect Dev tries*. 2007;1(3):289-95.
- [19] Christensen GD, Bisno AL, Simpson WA, Beachey EH. Adherence of slime producing strains of *Staphylococcus epidermidis* to smooth surfaces. *Infect Immun*. 1982;3(7):318-26.
- [20] Stepanovic S, Vukovi D, Hola V, et al. Quantification of biofilm in microtiter plates: overview of testing conditions and practical recommendations for assessment of biofilm production by *Staphylococci*. *APMIS*. 2007;11(5):891-99.
- [21] Amalaradjou MA, Narayanan A, Baskaran S, Venkitanarayanan K. Antibiofilm effect of trans-cinnamaldehyde on uropathogenic *Escherichia coli*. *J Urol*. 2010;18(4):358-63.
- [22] Vasilev K, Cook J, Griesser HJ. Antibacterial surfaces for biomedical devices. *Expert RevMed Devices*. 2009;6:553-67.
- [23] Costerton JW, Montanaro L, Arciola CR. Biofilm in implant infections: Its production and regulation. *Int J Artif Organs*. 2005;2(8):1062-68.
- [24] Trampuz A, Zimmerli W. New strategies for the treatment of infections associated with prosthetic joints. *Curr Opin Investig Drugs*. 2005;6:185-90.
- [25] Kaplan JB. Therapeutic potential of biofilm-dispersing enzymes. *Int J Artif Organs*. 2009;3(2):545-54.
- [26] Kaplan JB. Biofilm dispersal: mechanisms, clinical implications, and potential therapeutic uses. *J Dent Res*. 2010;8(9):205-18.
- [27] Gil ML, Casanova M, Martinez JP. Changes in the cell wall glycoprotein composition of *Candida albicans* associated to the inhibition of germ tube formation by EDTA. *Arch Microbiol*. 1994;16(1):489-94.
- [28] Root JL, McIntyre OR, Jacobs NJ, Daghlian CP. Inhibitory effect of disodium EDTA upon the growth of *Staphylococcus epidermidis* in vitro relation to infection prophylaxis of Hickman catheters. *Antimicrob Agents Chemother*. 1988;3(2):1627-31.
- [29] Venkatesh M, Rong L, Raad I, Versalovic J. Novel synergistic antibiofilm combinations for salvage of infected catheters. *Journal of Medical Microbiology*. 2009;5(8):936-44.
- [30] Chudzik B, Malm A, Rajtar B, Polz DM. In vitro activity of EDTA against planktonic and adherent cells of *Candida* sp. *Ann Microbiol*. 2007;5(7):115-19.
- [31] Marek J, Katarzyna P, Dariusz J, Anna M, Pawel R, Kazimierz G. EDTA as a Potential Agent Preventing Formation of *Staphylococcus Epidermidis* Biofilm On Polichloride Vinyl Biomaterials. *Ann Agric Environ Med*. 2008;1(5):237-41.
- [32] Amalaradjou MA, Venkitanarayanan K. Antibiofilm Effect of Otenidine Hydrochloride on *Staphylococcus aureus*, MRSA and VRSA. *Pathogens*. 2014; 3:404-16.
- [33] Percival SL, Kite P, Eastwood K, Murga R, Carr J, Arduino MJ. Tetrasodium EDTA as a novel central venous catheter lock solution against biofilm. *Infect Control Hosp Epidemiol*. 2005;2(6):515-19.
- [34] Yuk KK, Mendonca AF, Dong U. Effect of EDTA and Lysozyme on the Antimicrobial Activity of Ovotransferrin against *Listeria monocytogenes*. *Animal Industry Report*. 2010AS656, ASLR2495.
- [35] Young C, Otto M. *Staphylococcus epidermidis* infections. *Microbes Infect*. 2002;4:481-89.

### PARTICULARS OF CONTRIBUTORS:

1. PhD Scholar, Department of Microbiology, Medical College, Kolkata, West Bengal, India.
2. Professor and Head, Department of Microbiology, Medical College & Hospital, Kolkata, West-Bengal, India.
3. Professor, Department of Microbiology, Medical College, Kolkata, West Bengal, India.
4. Assistant Professor, Department of Microbiology, Medical College, Kolkata, India.
5. Assistant Professor, Department of Microbiology, Medical College, Kolkata, India.
6. PhD Student, IML, National Institute of Cholera and Enteric Diseases, (NICED) Kolkata, West Bengal, India.
7. PGT, Department of Microbiology, Medical College, Kolkata, West Bengal, India.

### NAME, ADDRESS, E-MAIL ID OF THE CORRESPONDING AUTHOR:

Dr. Manideepa Sen Gupta,  
Department of Microbiology, Medical College & Hospital, 88, College Street, Chitranjan Avenue, Kolkata, West-Bengal, India.  
E-mail : manideepa.sengupta2305@gmail.com

Date of Submission: **Nov 27, 2015**  
Date of Peer Review: **Dec 15, 2015**  
Date of Acceptance: **Dec 28, 2015**  
Date of Publishing: **Feb 01, 2016**

**FINANCIAL OR OTHER COMPETING INTERESTS:** None.